

High throughput screening of pKa by capillary electrophoresis and mass spectrometry (CE/MS) and long-term validation

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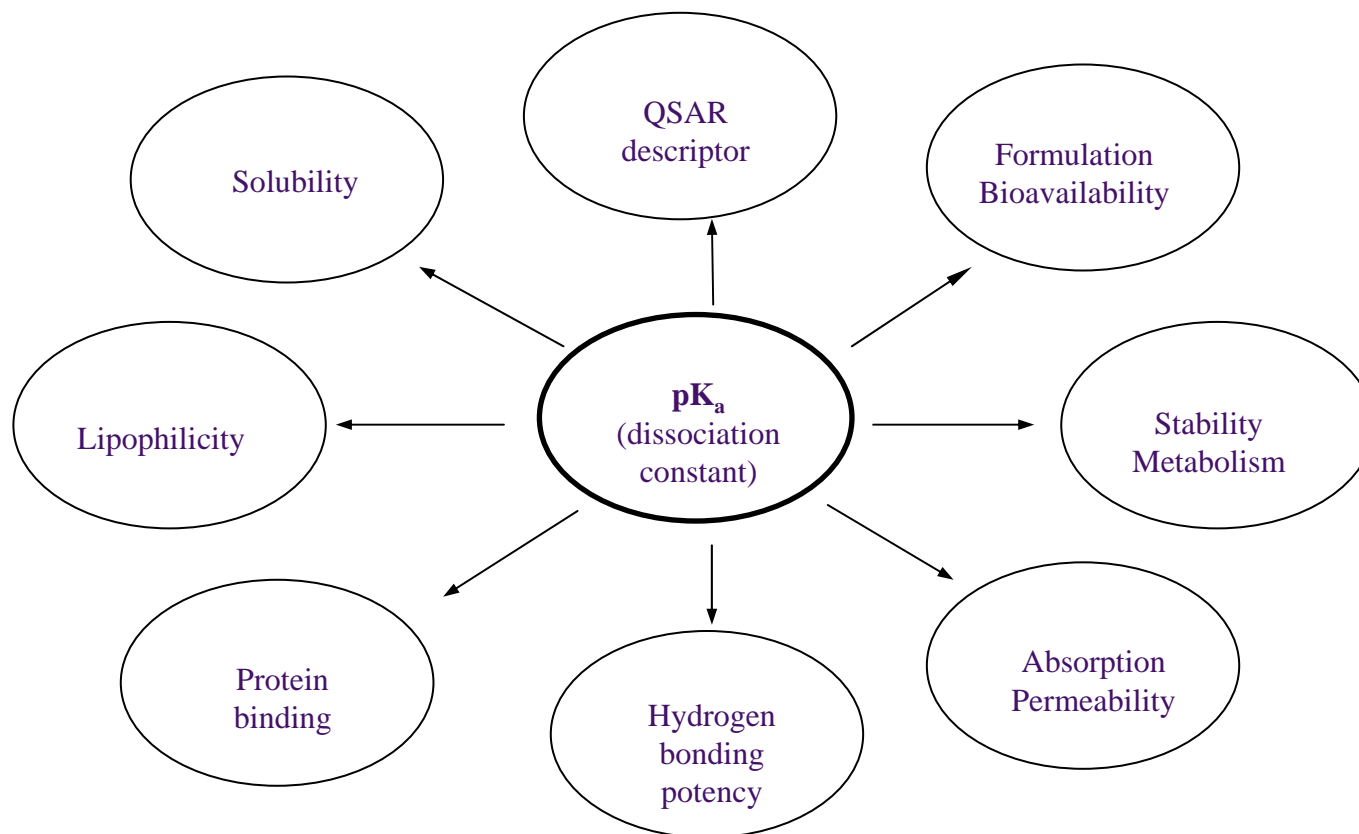
5th Physchem Forum, Stevenage, UK, June 19th

Outline

- Why screening pKa
- Advantages of CE/MS over established methods
- High throughput pKa screening method by CE/MS
 - Effects of buffer type and ionic strength on pKa
 - Comparison of CE/UV and CE/MS
 - Effects of pressure on migration times/sensitivity/mobility
- Comparisons of measured pKa and ref. and predicted values
- Reproducibility and accuracy and long-term validation
- Summary

Screening for pKa, why?

- Info about change in charge of molecule.....



Is CE/MS better technique for pKa than others?

- Sirius GLpKa – well established and widely used method !
- pION's Gemini for routine pKa (\$500/cdp, \$760/cpd/co-solvent)
- CE/UV (CombiSep), commercially available
- CE/MS (new technology), AZ, Mölndal, 2003 published.

H. Wan, A. Holmén, M. Någård, W. Lindberg, J. Chromatogr. A. 979 (2002) 369.

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Advantages of CE/MS over established methods

Specification	CE/MS	Titration/UV (D-PAS)
Amount of cpd	1 μg , or 1 μL of 10 mM solution	1-2 mg (titration) 3 μL of 10 mM DMSO stock (Sirius 3T)
Concentration	1-10 μM (100 μL)	> 20 μM (UV)
Purity required	No	Yes
Co-solvent	No	Yes, for poorly soluble cpds
Accuracy	< \pm 0.2	\pm 0.02-0.2?
Throughput capacity	>150 cpds/seq.(6h)	4 min/cpd (Sirius T3)
Limitation	Poor ionization (ESI)	Impurity? poor solubility (precipitation), Ionization group close to chromophore

Principle of pKa determination by CE

pKa \leftrightarrow 50% ionization

$$K_a^{\text{th}} = \frac{\{\text{H}^+\}\{\text{A}^-\}}{\{\text{HA}\}}$$

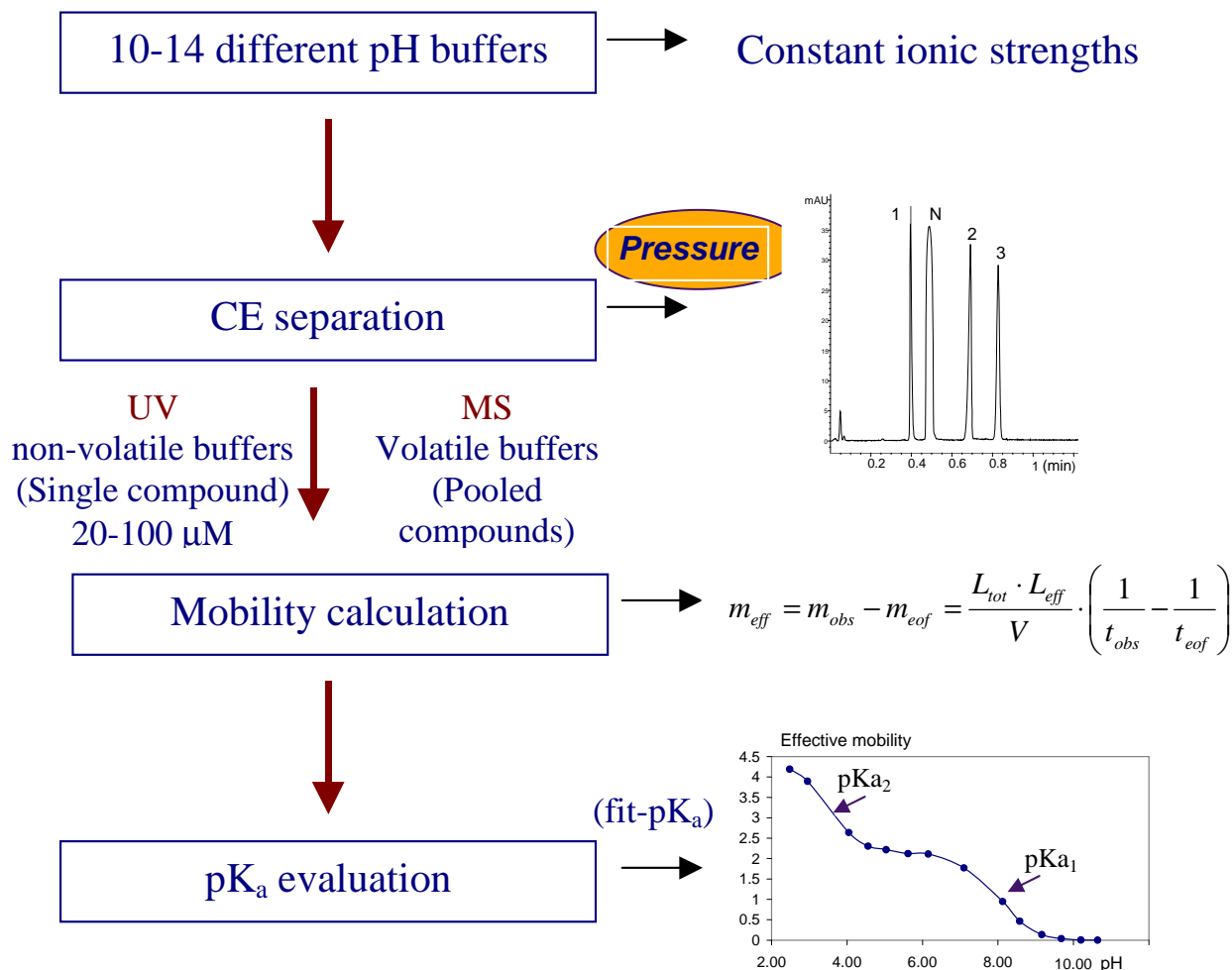
$$\text{pK}_a^{\text{th}} = \text{pH} - \log \left[\frac{m_{\text{eff}}}{m_m - m_{\text{eff}}} \right] + \frac{0.5085z^2\sqrt{I}}{1 + 0.328\alpha\sqrt{I}}$$

Equations used for pKa calculation

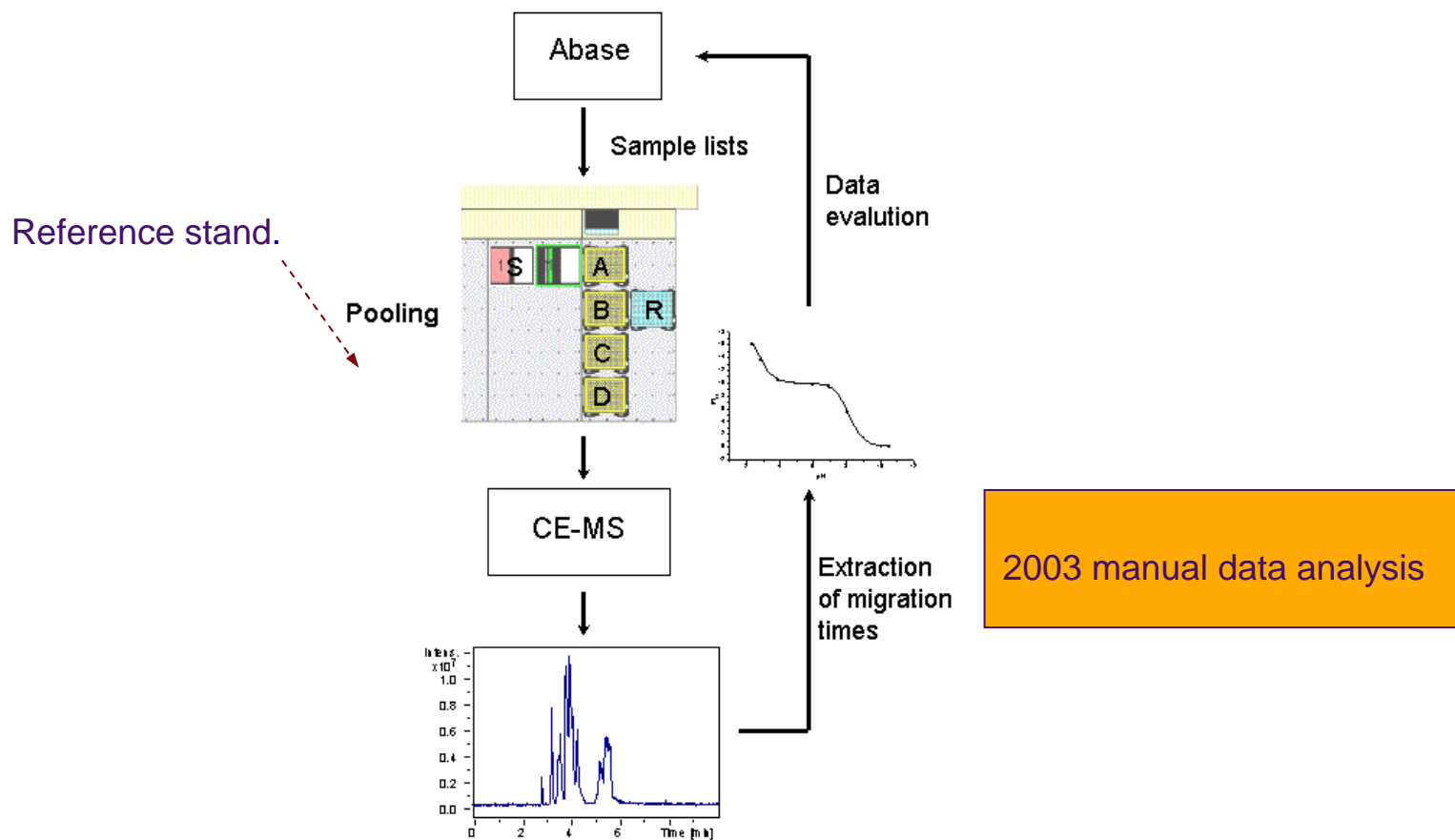
Ionizable type	Model equation
Monobase	$m_{\text{eff}} = \frac{M_b 10^{-\text{pH}}}{10^{-\text{pK}_a} + 10^{-\text{pH}}}$
Monoacid	$m_{\text{eff}} = \frac{M_a 10^{-\text{pK}_a}}{10^{-\text{pK}_a} + 10^{-\text{pH}}}$
Dibase	$m_{\text{eff}} = \frac{M_{b2} [10^{-\text{pH}}]^2 + M_{b1} 10^{-\text{pK}_{a1}} 10^{-\text{pH}}}{[10^{-\text{pH}}]^2 + 10^{-\text{pK}_{a1}} 10^{-\text{pH}} + 10^{-\text{pK}_{a1}} 10^{-\text{pK}_{a2}}}$
Diacid	$m_{\text{eff}} = \frac{M_{a1} 10^{-\text{pK}_{a1}} 10^{-\text{pH}} + M_{a2} 10^{-\text{pK}_{a1}} 10^{-\text{pK}_{a2}}}{[10^{-\text{pH}}]^2 + 10^{-\text{pK}_{a1}} 10^{-\text{pH}} + 10^{-\text{pK}_{a1}} 10^{-\text{pK}_{a2}}}$
Monoacidic monobasic ampholyte	$m_{\text{eff}} = \frac{M_{b1} [10^{-\text{pH}}]^2 + M_{a1} 10^{-\text{pK}_{a1}} 10^{-\text{pK}_{a2}}}{[10^{-\text{pH}}]^2 + 10^{-\text{pK}_{a1}} 10^{-\text{pH}} + 10^{-\text{pK}_{a1}} 10^{-\text{pK}_{a2}}}$
Tribase	$m_{\text{eff}} = \frac{M_{b3} [10^{-\text{pH}}]^3 + M_{b2} 10^{-\text{pK}_{a1}} [10^{-\text{pH}}]^2 + M_{b1} 10^{-\text{pK}_{a1}} 10^{-\text{pK}_{a2}} 10^{-\text{pH}}}{[10^{-\text{pH}}]^3 + 10^{-\text{pK}_{a1}} [10^{-\text{pH}}]^2 + 10^{-\text{pK}_{a1}} 10^{-\text{pK}_{a2}} 10^{-\text{pH}} + 10^{-\text{pK}_{a1}} 10^{-\text{pK}_{a2}} 10^{-\text{pK}_{a3}}}$
Triacid	$m_{\text{eff}} = \frac{M_{a1} 10^{-\text{pK}_{a1}} [10^{-\text{pH}}]^2 + M_{a2} 10^{-\text{pK}_{a1}} 10^{-\text{pK}_{a2}} 10^{-\text{pH}} + M_{a3} 10^{-\text{pK}_{a1}} 10^{-\text{pK}_{a2}} 10^{-\text{pK}_{a3}}}{[10^{-\text{pH}}]^3 + 10^{-\text{pK}_{a1}} [10^{-\text{pH}}]^2 + 10^{-\text{pK}_{a1}} 10^{-\text{pK}_{a2}} 10^{-\text{pH}} + 10^{-\text{pK}_{a1}} 10^{-\text{pK}_{a2}} 10^{-\text{pK}_{a3}}}$
Diacidic monobasic ampholyte	$m_{\text{eff}} = \frac{M_{b1} [10^{-\text{pH}}]^3 + M_{a1} 10^{-\text{pK}_{a1}} 10^{-\text{pK}_{a2}} 10^{-\text{pH}} + M_{a2} 10^{-\text{pK}_{a1}} 10^{-\text{pK}_{a2}} 10^{-\text{pK}_{a3}}}{[10^{-\text{pH}}]^3 + 10^{-\text{pK}_{a1}} [10^{-\text{pH}}]^2 + 10^{-\text{pK}_{a1}} 10^{-\text{pK}_{a2}} 10^{-\text{pH}} + 10^{-\text{pK}_{a1}} 10^{-\text{pK}_{a2}} 10^{-\text{pK}_{a3}}}$
Monoacidic dibasic ampholyte	$m_{\text{eff}} = \frac{M_{b2} [10^{-\text{pH}}]^3 + M_{b1} 10^{-\text{pK}_{a1}} [10^{-\text{pH}}]^2 + M_{a1} 10^{-\text{pK}_{a1}} 10^{-\text{pK}_{a2}} 10^{-\text{pK}_{a3}}}{[10^{-\text{pH}}]^3 + 10^{-\text{pK}_{a1}} [10^{-\text{pH}}]^2 + 10^{-\text{pK}_{a1}} 10^{-\text{pK}_{a2}} 10^{-\text{pH}} + 10^{-\text{pK}_{a1}} 10^{-\text{pK}_{a2}} 10^{-\text{pK}_{a3}}}$

Graphs and table from Miller, *Electrophoresis* 2002, 23, 2833

High-throughput pKa screening by CE/MS

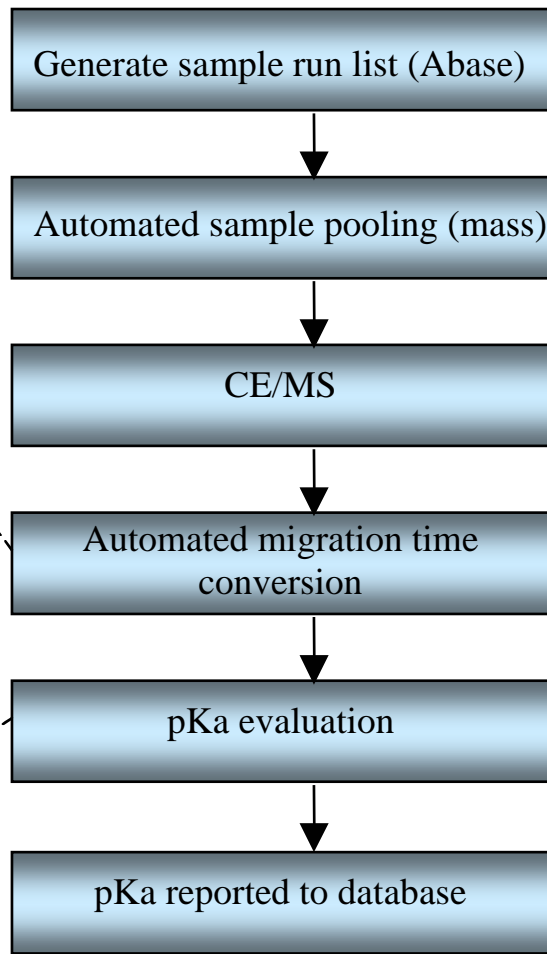
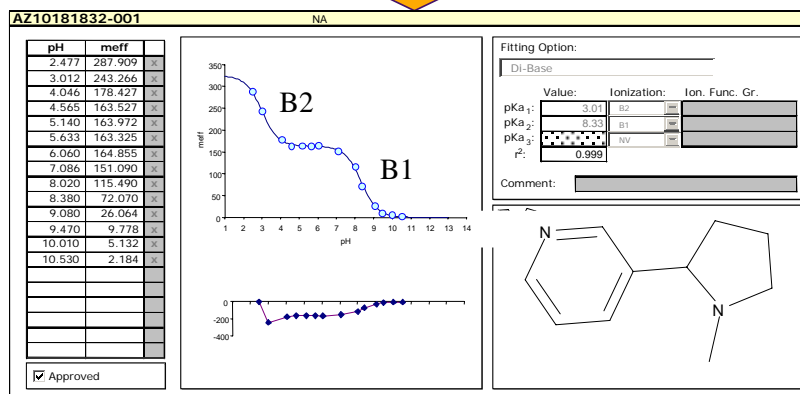
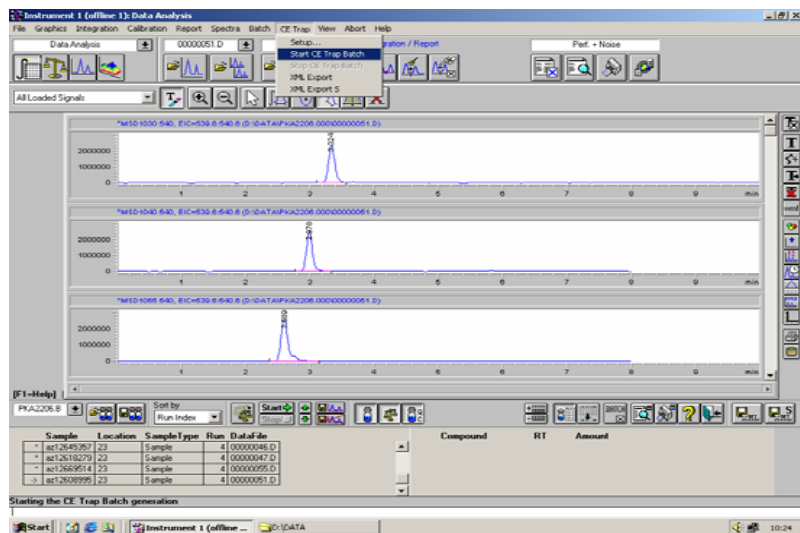


Sample pooling and data analysis flow scheme

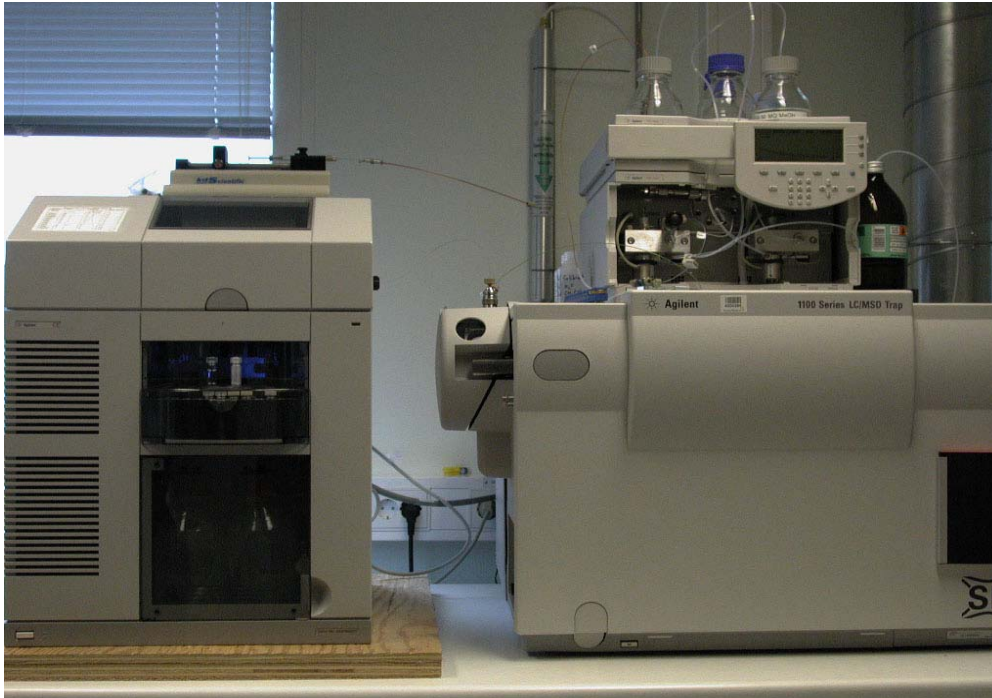


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Rapid Commun. Mass Spectrom. 17 (2003) 2639.

Current pKa screening - integrated & automated assay



CE/MS instrumentation

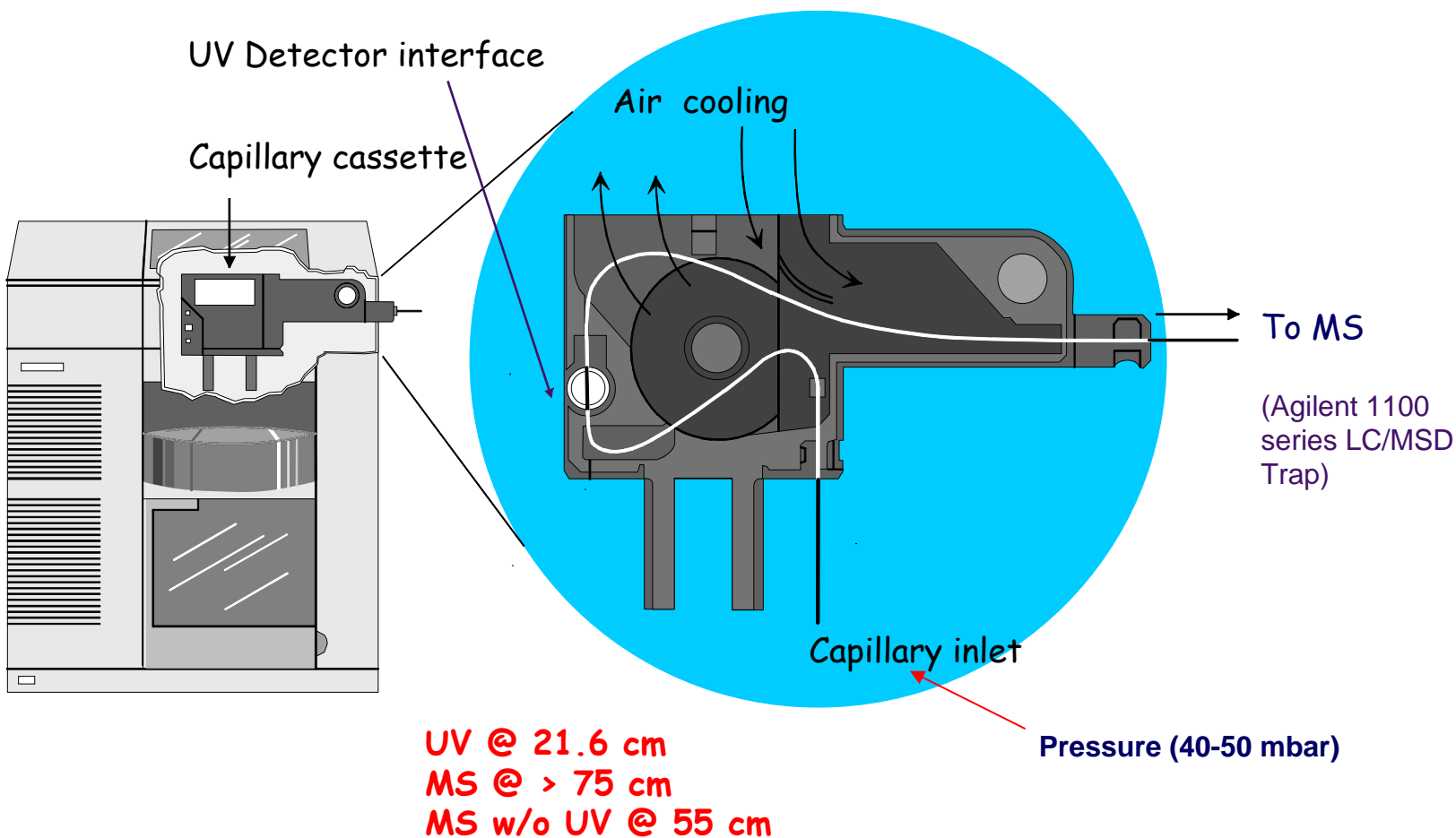


- Higher initial investment
- Long-term savings

HPCE^{3D} , 1100 series LC/MSD trap SL (Agilent)

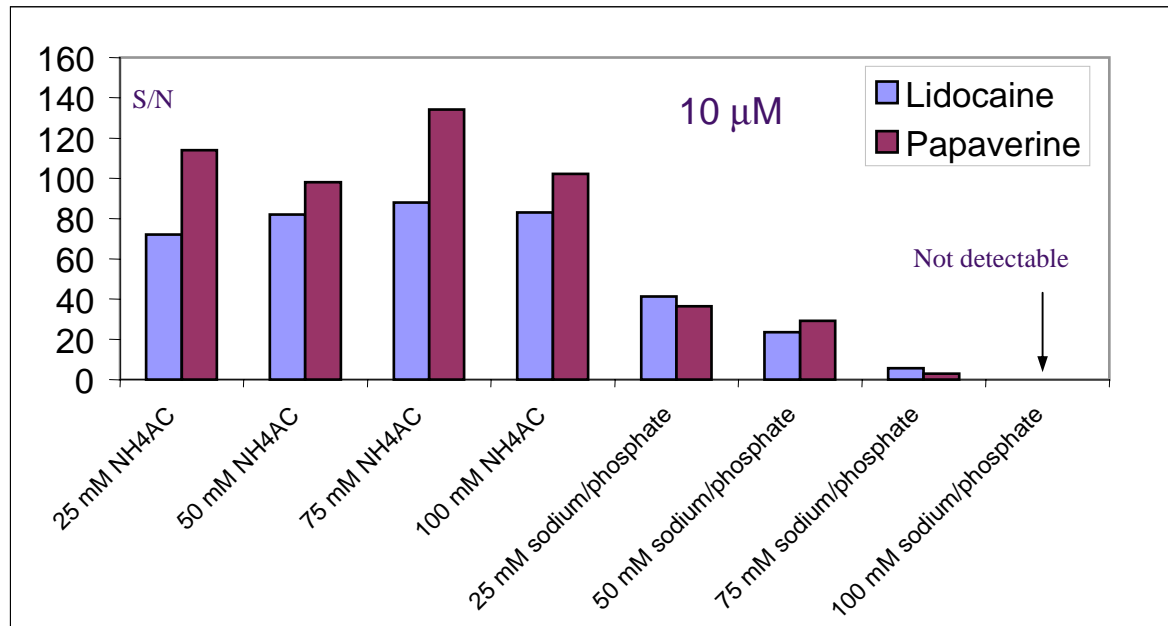
- Untreated fused silica capillary (50-60 cm/50 SEK) can be used as long as it works.
- Buffers without filtration (stock solutions can be used up to >3 years).
- Sheath liquid (1 L) can be used for more than 6 months (99% recycling).

On-line CE/UV/MS experimental setup



Courtesy from Agilent (with small modifications)

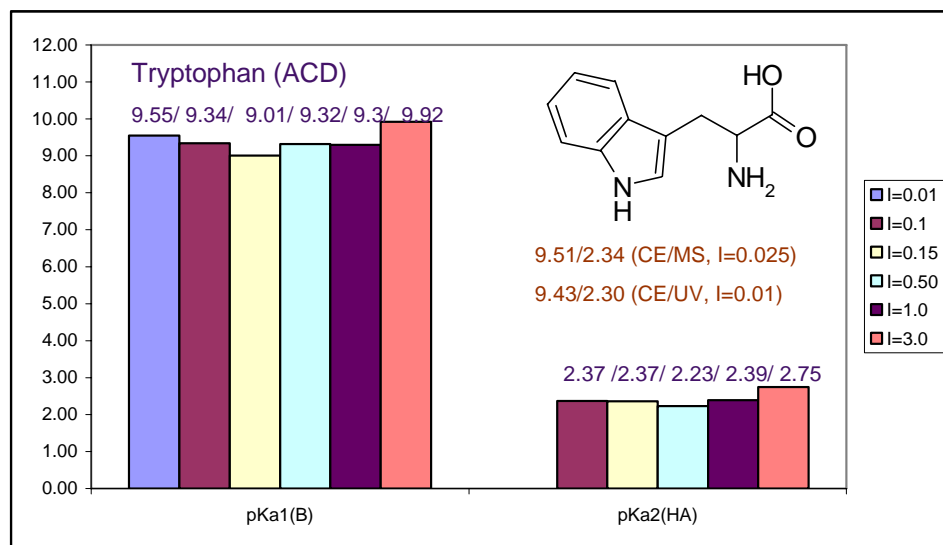
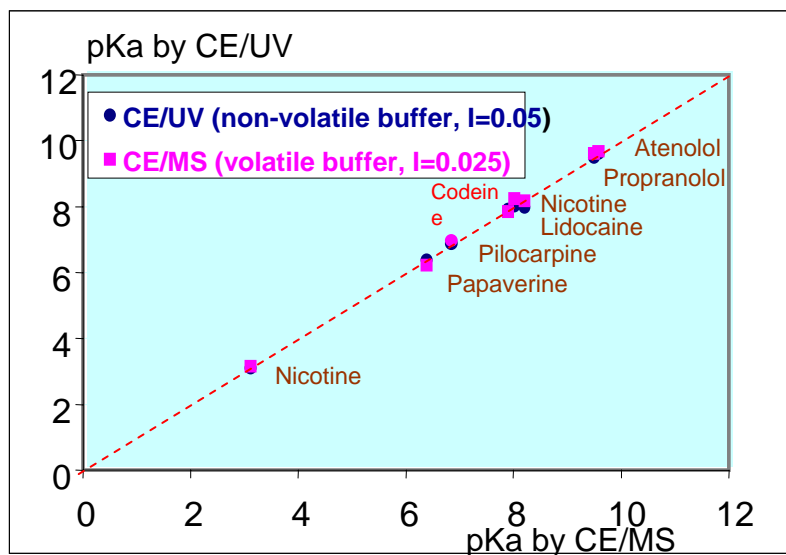
Sensitivity from volatile vs non-volatile buffers in CE/MS



- Non-volatile buffer decreased signals at higher concentration (ion suppression)
- Ion source contamination by non-volatile buffer

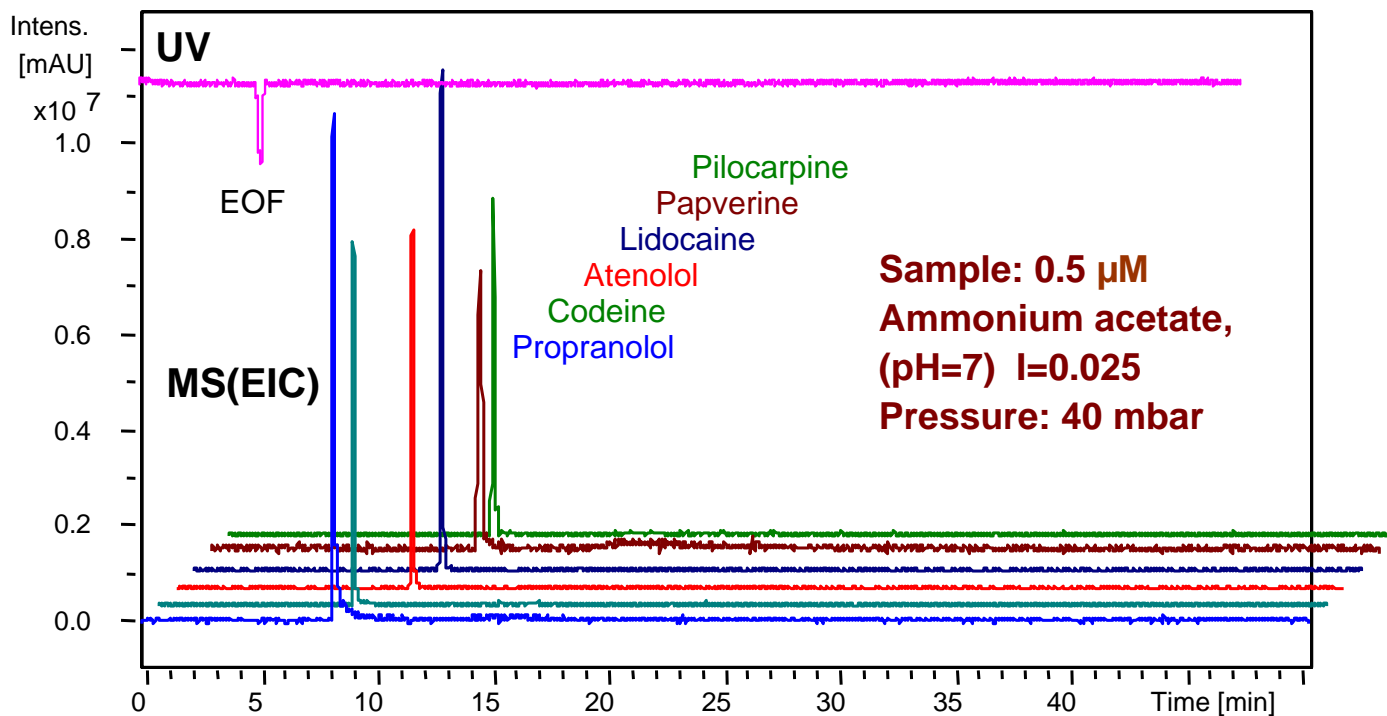
Effects of buffer type and ionic strength on pKa

$$pK_a^{th} = pH - \log \left[\frac{m_{eff}}{m_m - m_{eff}} \right] + \frac{0.5085z^2 \sqrt{I}}{1 + 0.328\alpha\sqrt{I}}$$



- Titration method uses physiological buffer with ionic strength at 0.15 M.
- Ionic strengths (0.025 - 0.15 M) have a small effect on pKa ($\Delta pK_a=0.064$)

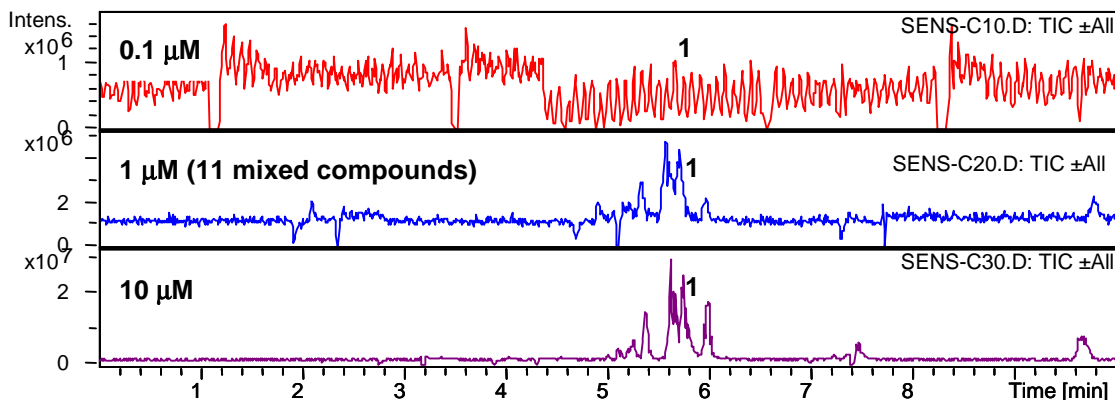
Comparison of UV and MS (Ion trap) sensitivity



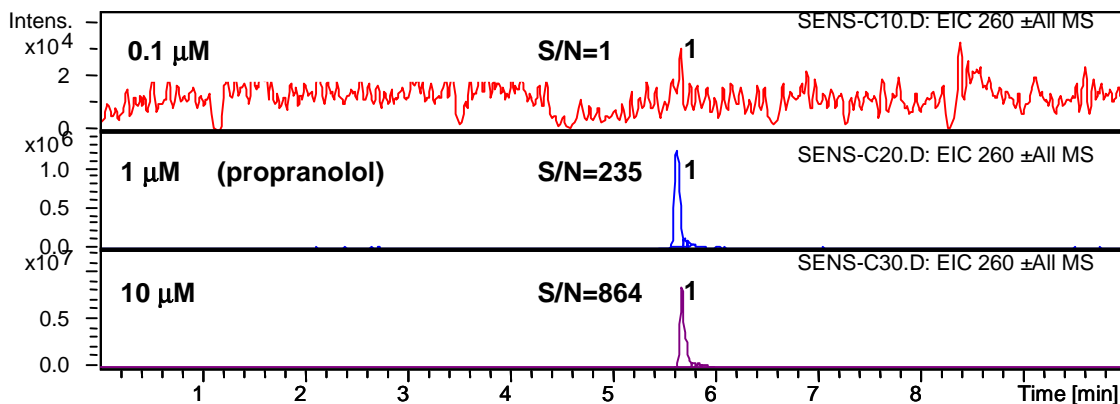
UV working concentration: 20-100 μ M

Sensitivity and reproducibility of pressure-assisted-CE/MS

Total ion chromatogram (TIC)

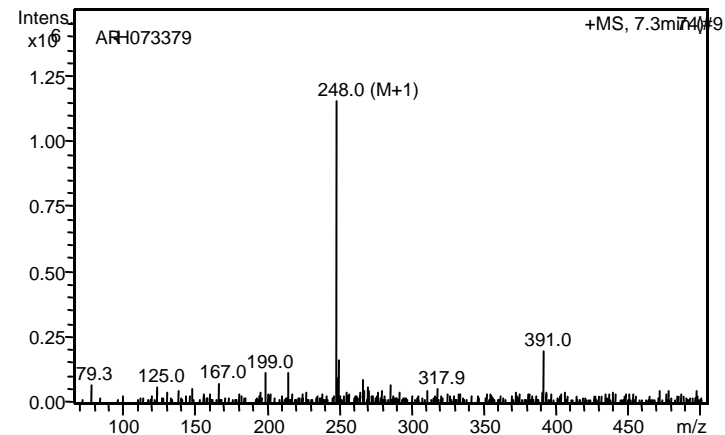
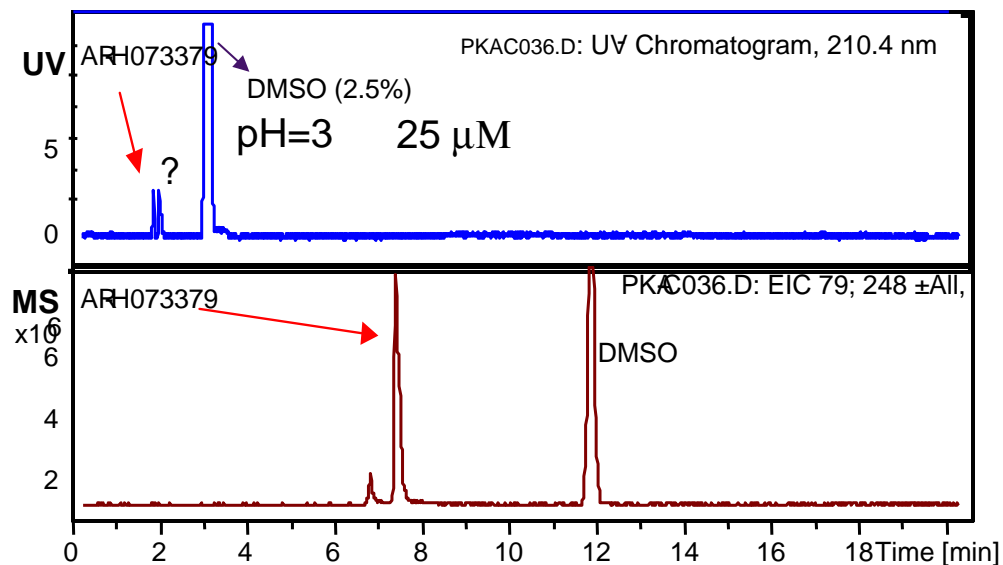


(EIC) and migration times after 30 runs



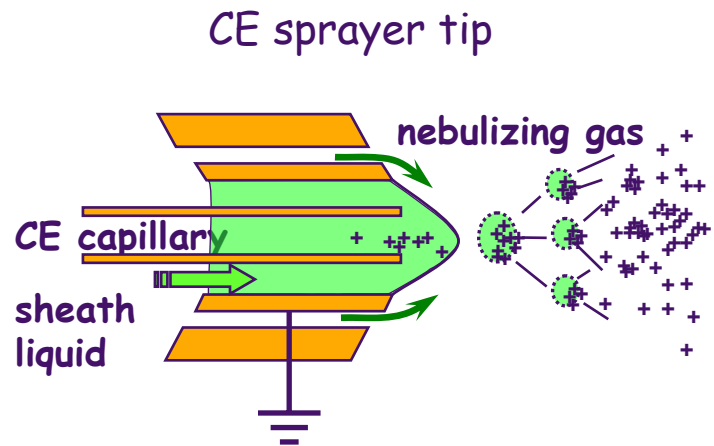
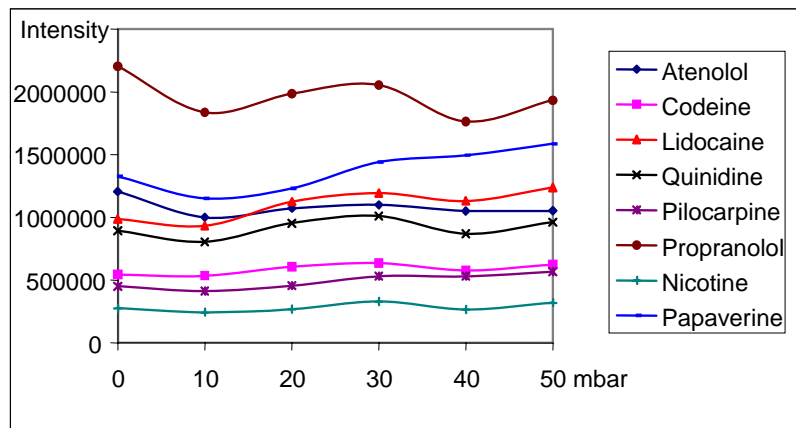
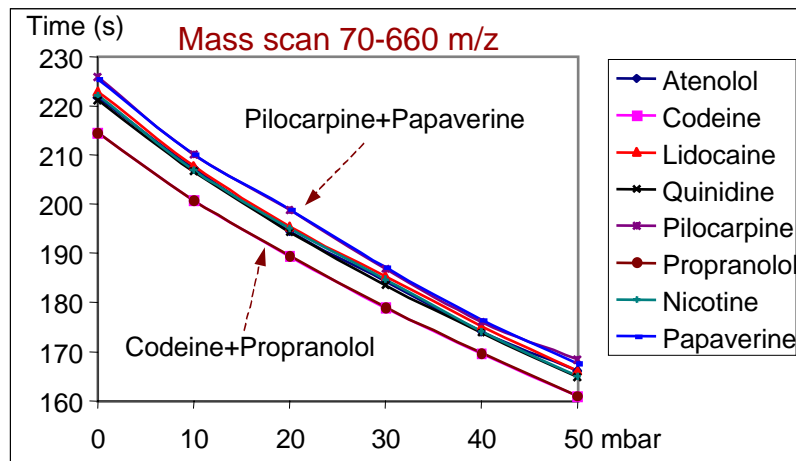
**Ammonium acetate
(pH=7) I=0.025;
pressure, 40 mbar**

Sensitive and selective MS detection over UV



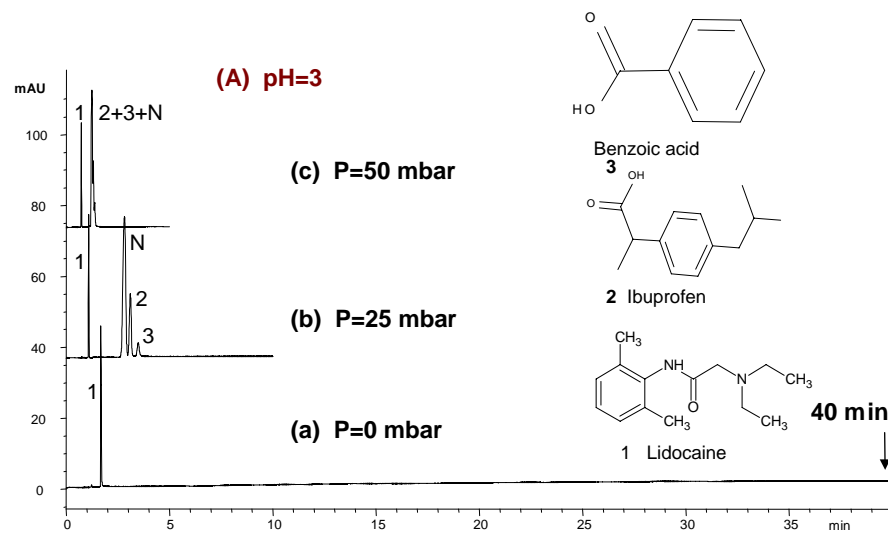
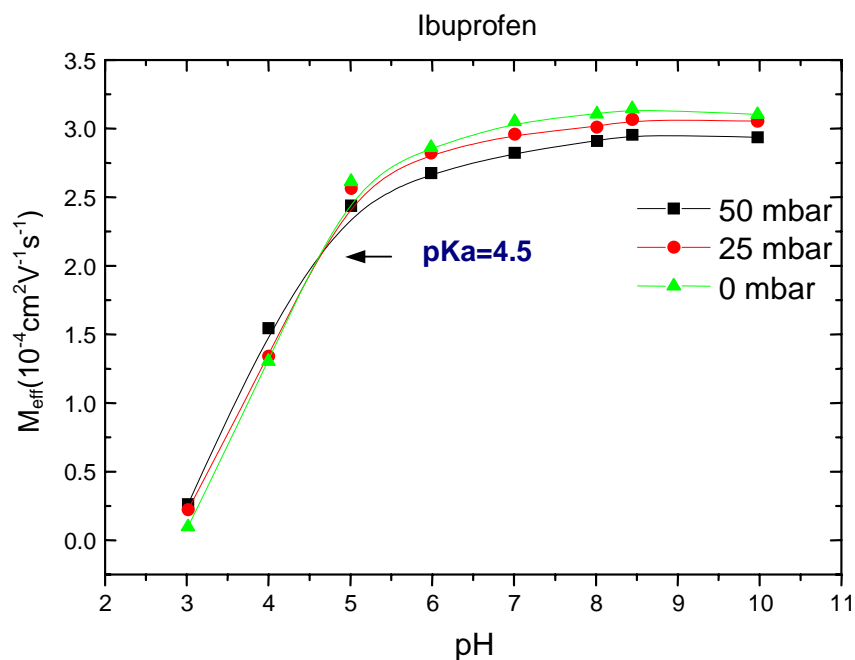
On-line tandem UV and MS detection

Effects of pressure on migration times/sensitivity



- Pressure reduced migration times
- Negligible signal suppression
- Improved RSD of effective mobility
- High throughput capacity

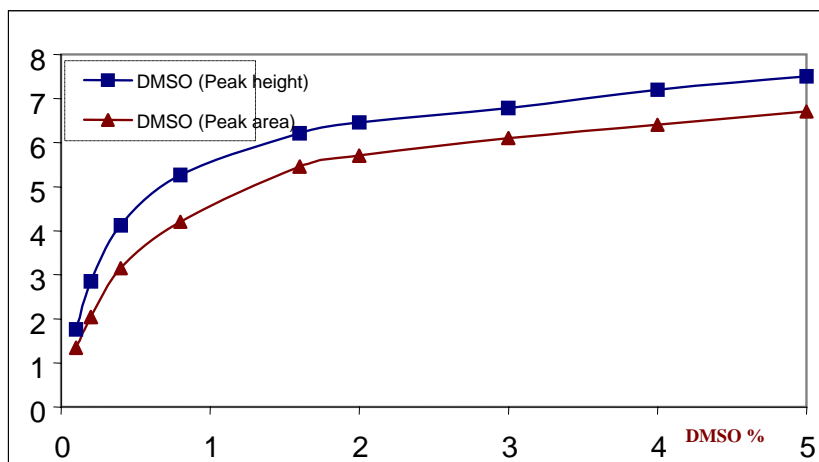
Effects of pressure on effective mobility



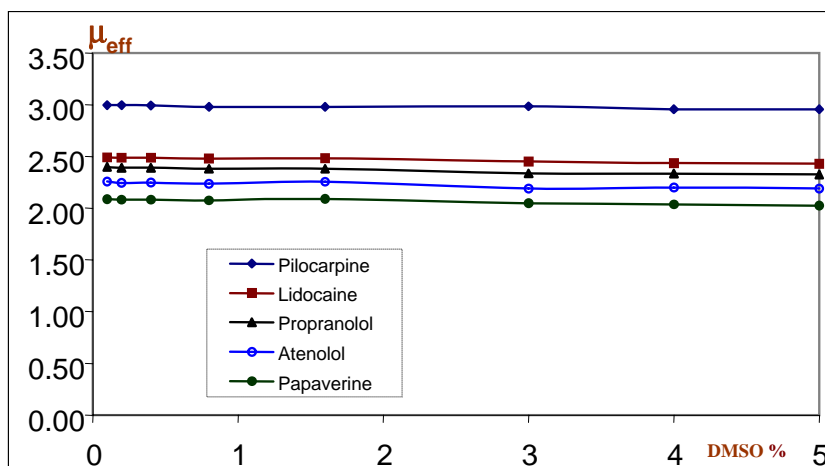
- Effective mobility shift around pKa (an interesting observation)
- Mobility shift caused by pressure doesn't affect pKa values.

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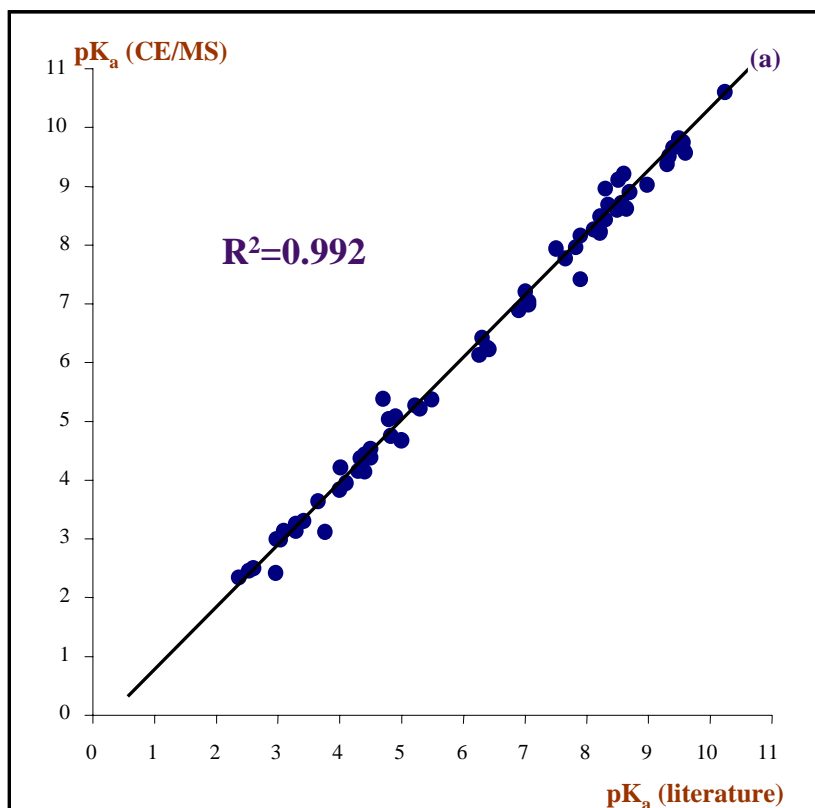
How many compounds can be pooled by CE/MS?



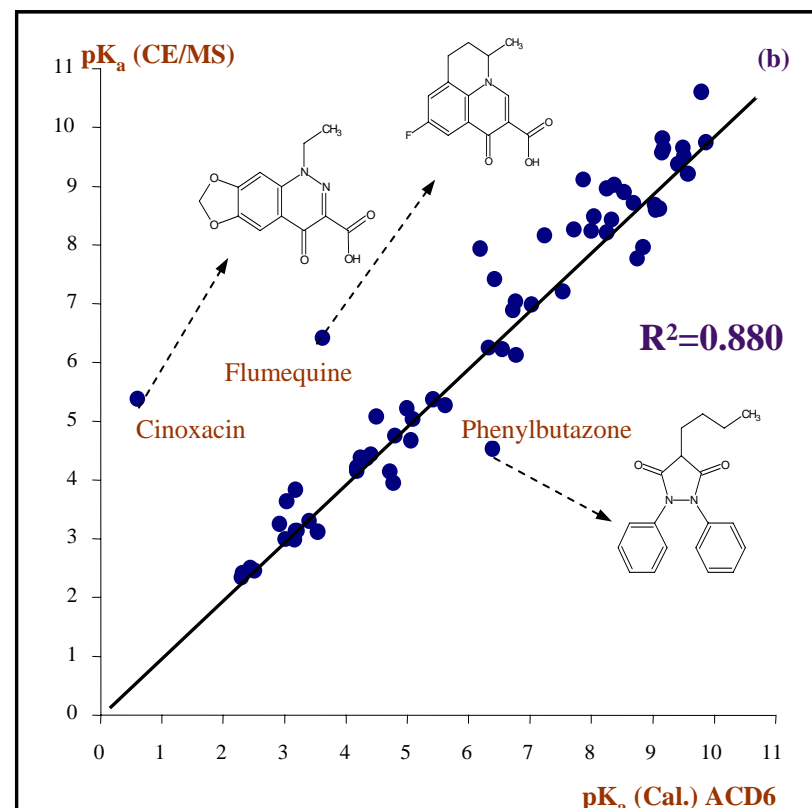
- DMSO < 5% in sample
- Constant mobility
- High MS resolution
- More than 150 cpds/sequence



Comparison of measured pKa and lit./predicted values

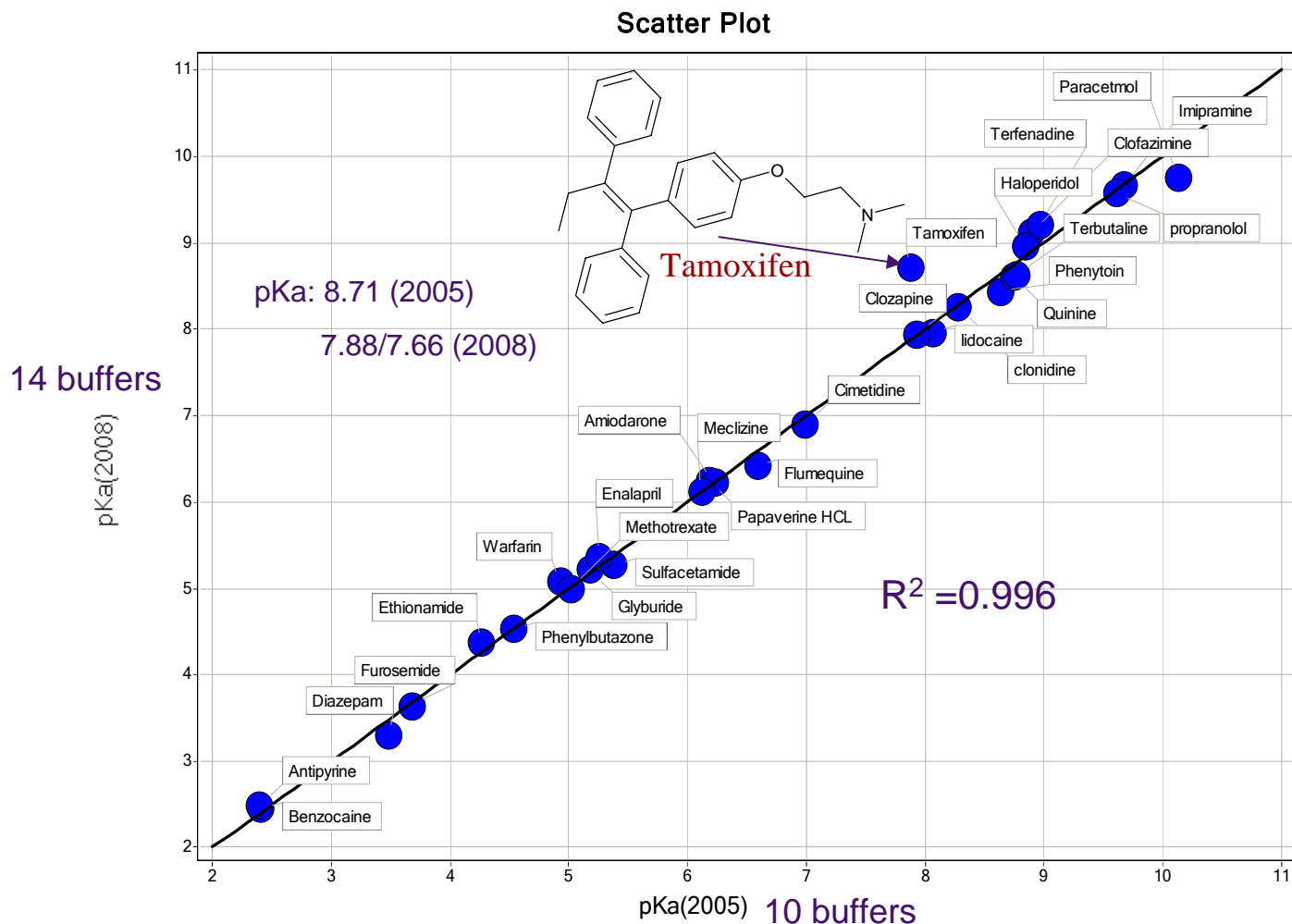


Poor prediction for conjugated structures

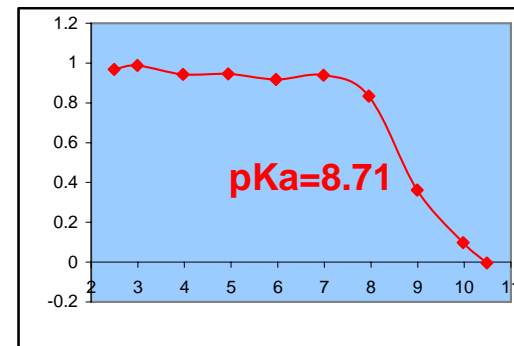
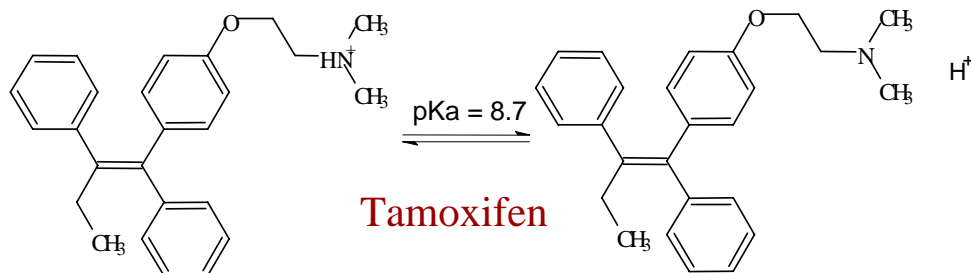


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Long-term validation of CE/MS pKa (2005-2008)

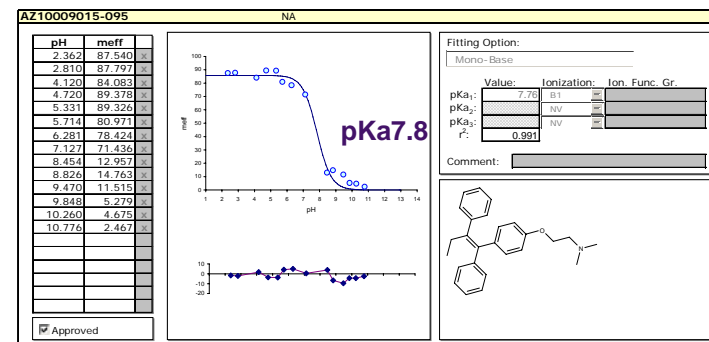


An example of poorly soluble compound



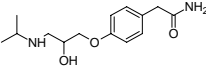
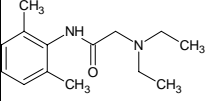
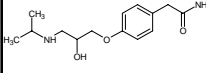
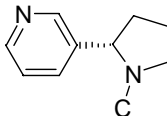
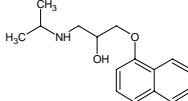
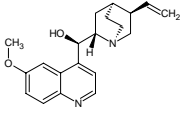
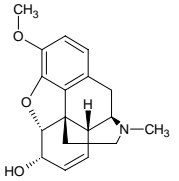
1. Ionization not close to chromophore
2. Low solubility 0.45 μM (pH 7.4), LogD=6.5

pKa (DPAS): no
 pKa (GlpKa): co-solvent, precipitation ?
 pKa (CE/MS): 8.71 (2005), 7.88/7.76 (2008)
 (ref: 7.6)



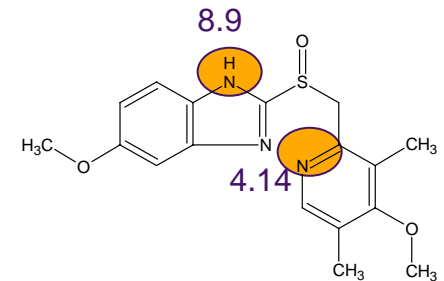
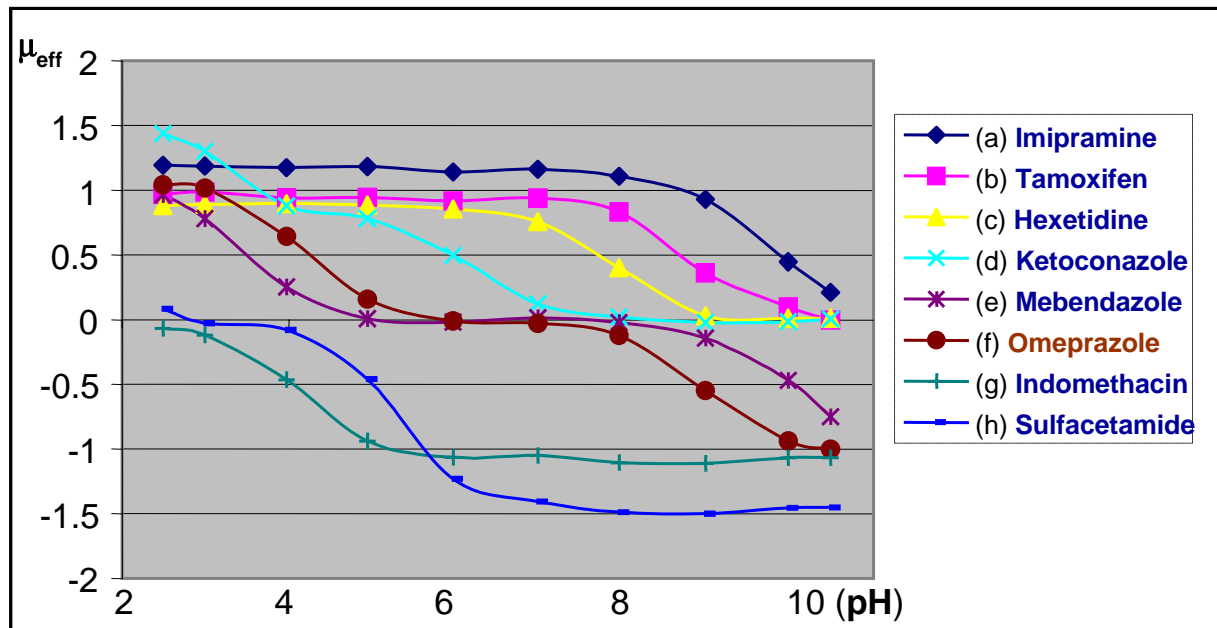
Ref. (Karl Box et al., Anal. Chem., 2003, 75, 883-892).

Long-term reproducibility of QC (2005-2008)

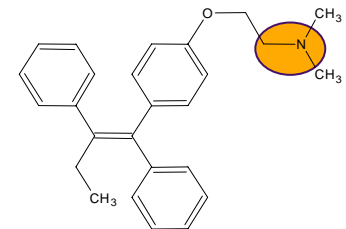
QC	AZ_1	AZ_2	AZ_3	AZ_4	AZ_5	AZ_6	AZ_7
Compound	Atenolol	Lidocaine	Papaverine	Nicotine	Propranolol	Quinine	Codeine
Structure							
pKa(B1/B2) Mean (n=50)	9.73	8.16	6.05	8.45(3.12)	9.67	8.88(4.00)	8.45
SD	0.09	0.09	0.11	0.10(0.27)	0.08	0.10(0.19)	0.08
Ref.	9.58	7.90	6.39/5.95	8.12(3.12)	9.50	8.50(4.1) 8.90 8.54	8.21 8.30
CE/UV	9.61	7.92	6.38	8.02(3.10)	9.49	8.39(4.14)	7.97

- Seven compounds as on-line QC for pKa screening (single measurement)
- Reproducibility <0.2 pKa units

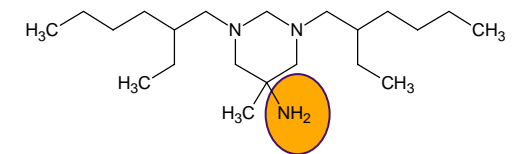
Examples of determination of poorly soluble compounds



(f) Omeprazole



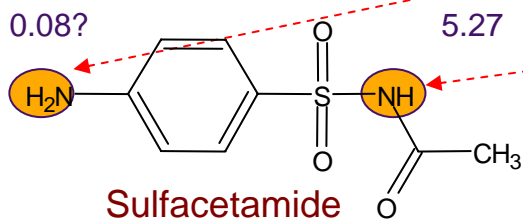
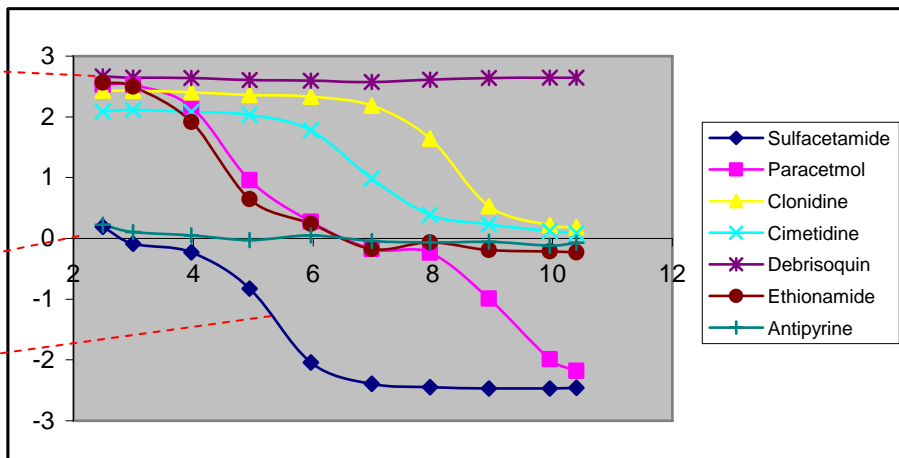
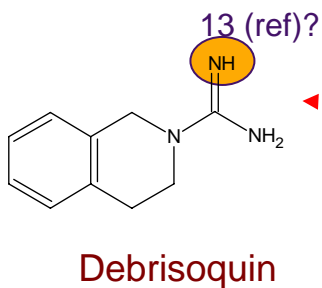
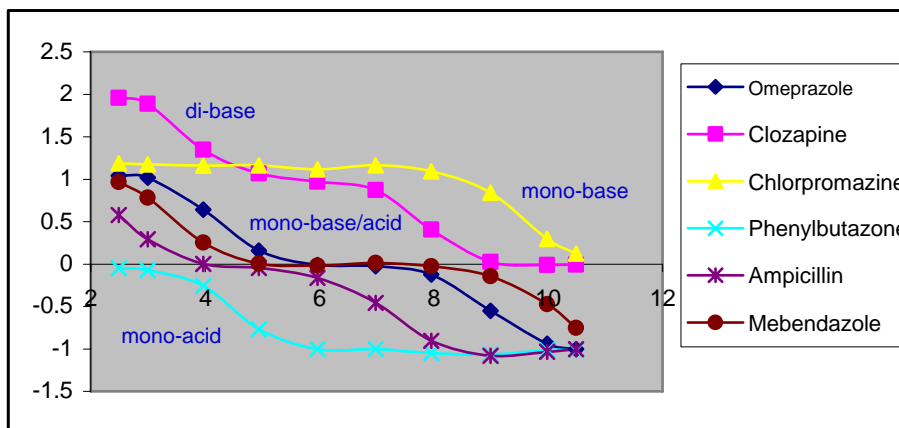
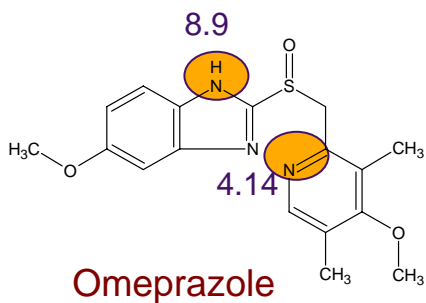
(b) Tamoxifen



(c) Hexetidine

- Possible to measure pKa with solubility < 1 μM
- $\text{pKa} = 50\% \text{ ionization} \leftrightarrow (\text{max. mobility})/2$
- Mobility \approx charge/mass

pKa fitting example pH 2.5-10.5/10 buffers(2.3-10.8/14 buffers)



Summary

- Pressure-assisted CE/MS for pKa:
 - High throughput screening pKa from 2.3 to 10.8
 - Reproducibility and accuracy of pKa \pm 0.2 units
 - Insensitive to compound purity, requiring minute sample
 - Concn: (1-10 μ M) beneficial for poorly soluble cpds
 - Independent on type of buffer, capillary length, DMSO concn.
 - Provide charge and structure/stability information (MS)

Acknowledgements

- Anders Holmén, Mats Någård, Walter Lindberg, Yudong Wang, Marie Englund, Richard Thompson, Johan Ulander

H. Wan, A. Holmén, M. Någård, W. Lindberg, J. Chromatogr. A. 979 (2002) 369.

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H. Wan, R. Thompson; Drug Discovery Today, Technologies, 2(2005), 171.

H. Wan, J. Ulander, Expert Opinion on Drug Metabolism & Toxicology, 2 (2006), 1389.

- Eva Emanuelsson (validation), Fredrik Bergström (new pKa-fit template test)
- Eva Thorin (Activitybase/Dixy group), Michael Wirth Färdigh (automation)
- Zhiping You (new pKa-fit program), AZ Boston
- Lead generation, Physical Chemistry group, AZ Mölndal